AMENDMENTS TO THE SPECIFICATION

Please amend the specification as follows:

Please replace the first paragraph starting on page 23, line 3 with the following rewritten paragraph:

Total RNA was reverse-transcribed (RT) into cDNA and polymerase chain reaction (PCR) was performed in the same reaction using a real-time ~~TAQman~~ TAQMAN® One-Step RT-PCR Master Mix Reagents Kit (obtained from Applied Biosystems; Foster City, Calif.). The sequences of the primer/probe sets used for this analysis are as follows. F and R are the forward and reverse primers, respectively, and P is the fluorescent-labeled probe.

Please replace the paragraph starting on page 23, line 19 with the following rewritten paragraph:

The real-time one step RT-PCR cycling conditions for all primer sets were as follows: 30 min at 48 °C for RT step; 10 min at 95° C for ~~AMPLITAQ Gold~~ AMPLITAQ GOLD® DNA polymerase Activation; and 40 cycles for cDNA denaturing (95° C, 15 s), and annealing/elongation (60° C for 1 min) steps. PCR reactions for each template were done in triplicate using 1 µg of total RNA per sample. Each gene-specific primer pair was tested on standard 384-well plates. Standard curves were constructed using 10-1000 ng of total RNA prepared from the CWRSA6 tumor line. All experiments were optimized such that the threshold cycle (C_T) from triplicate reactions did not differ by more than one cycle number.

Please replace the paragraph starting on page 24, line 33 with the following rewritten paragraph:

Raw 267.4 cells were maintained at 37° C. and 5% CO_2 in Dulbecco's modified eagle medium (DMEM) with high glucose (obtained from ~~GIBCO~~ GIBCO®; Grand

Island, N.Y.) supplemented with 10% fetal bovine serum (FBS), 100 µg/ml penicillin and 100 µg/ml streptomycin.

Please replace the paragraph starting on page 25, line 28 with the following rewritten paragraph:

Prostate carcinoma cell lines, 22Rv1 and LNCaP, and human acute monocytic leukemia (THP-1) cells were obtained from the American Type Culture Collection (Rockville, Md.) and maintained in RPMI 1640 medium supplemented with 10% FBS, 100 µg/ml penicillin and 100 µg/ml streptomycin (obtained from ~~GIBCO~~ GIBCO®; Grand Island, N.Y.). Three different assays demonstrate that R-etodolac has the ability to function as a PPARγ transactivator.

Please replace the paragraph starting on page 26, line 31 with the following rewritten paragraph:

Third, PPARγ is accepted as a master regulator of adipocyte differentiation. Uptake of neutral lipids is a marker of adipocyte differentiation and Oil Red O staining of these neutral lipids is an accepted procedure to demonstrate this differentiation phenomenon (Tontonoz *et al.*, 1998). NIH3T3 cells that stably overexpress retrovirally expressed recombinant PPARγ were obtained from Dr. Ronald Evans (Salk Institute, La Jolla, Calif.). Cells treated with 1 µM R-etodolac displayed accumulation of neutral lipids and morphological changes associated with PPARγ activity that are comparable to those observed with troglitazone at a similar concentration (Figure 1C). NIH3T3 cells stably expressing recombinant PPARγ (obtained from Dr. Ronald Evans of the Jonas Salk Institute; San Diego, Calif.) were maintained in DMEM supplemented with 10% BCS, 100 µg/ml penicillin and 100 µg/ml streptomycin (obtained from ~~GIBCO~~ GIBCO®; Grand Island, N.Y.). The cells were treated for seven days with the indicated compounds and concentrations and stained for neutral lipids with Oil Red O as described by Green and Kehinde ("Sublines of mouse 3T3 cells that accumulate lipid," Cell, Vol. 1, p. 113-116 (1974)). The lipid uptake was dose-dependent and was significantly more pronounced at 500 µM concentration of R-etodolac. NIH3T3 cells transfected with the empty retroviral vector did not demonstrate the lipid uptake with

either troglitazone or R-etodolac (data not shown). Having demonstrated that PPARγ could be positively modulated by R-etodolac, the possibility of using it as a potential therapeutic against prostate tumor models was considered.